

Antioxidant Activity of Bacterial Cellulose Based Edible Films Incorporated with Catechin and *Citrus aurantium* L (Bergamot) Essential Oil

Yoice Srikandace^{1*}, Sylviana², Nina Artanti³, Zalinar Udin³ and M.Hanafi³

¹Indonesian Institute of Sciences-Research Unit for Clean Technology,
Jl. Cicitu-Sangkuriang Bandung, Indonesia.

²Departement of Nutrition and Food Technology, Faculty of Life Sciences, Surya University
SETOS Building, Jl. M.H. Thamrin Km. 2.7, Tangerang, Indonesia

³Indonesian Institute of Sciences-Research Center for Chemistry,
Jl. Kw.Puspiptek, Muncul-Serpong, Tangerang, Indonesia

*E-mail: yoice.srikandace.s@gmail.com

Abstract: Bacterial cellulose-based edible films have been known as natural material and eco-friendly fibers for food packaging. The edible films contain carboxymethyl cellulose (CMC), Tween 80 and glycerol as an external food protection. The edible films as carriers of ingredients agents (antimicrobial, antioxidant) preserve the quality of food product. Incorporating antioxidant compounds into edible films provide the novel approach to improve the safety, shelf-life of foods and physical properties. In present study, the antioxidant agents are catechin and bergamot oil. The study was aimed to investigate the antioxidant activities of edible films incorporated with catechin (FC), bergamot oil (FB) and both catechin and bergamot oil (FCB). Antioxidant activity was carried out using DPPH method. The results showed that antioxidant activities of FC, FB and FCB with various concentrations antioxidant agents (1-2.5%) were 65.78%, 73.16% and 81.07% respectively. GC-MS showed bergamot oil contained compounds limonene (23,66%), linalool (19,44), and linalil acetate (37,88). SEM showed the surface of edible films FB and FCB were fine and smooth. Edible film only and FC revealed the cracks on the surfaces of films. All compounds of edible films were still available based on FTIR results.

Key words: *Bacterial cellulose, Bergamot oil, Catechin, Edible film, Packaging*

INTRODUCTION

Bacterial cellulose (biocellulose) is pure extracellular fiber produced by *Acetobacter spp* that has numerous applications in food, biomedical and paper industries [1]. Biocellulose was produced by *Gluconacetobacter xylinum* (formerly *Acetobacter xylinum*) using coconut water and it is popularly known as nata-de-coco [2][3]. In general, the production of nata de coco is done by direct inoculation into liquid medium. The average time for producing nata was 11 days, with an average thickness of 0.8 cm [3]. Bacterial cellulose fibers exhibited the very pure fiber, very stiffness, strength, high thermal stability and similar to plant cellulose [4][5]. It also displays unique properties including high tensile strength, high water absorption capacity, high crystallinity and an ultra-fine[4]. Nowadays, bacterial cellulose also provides biodegradable material package in order to reduce the use of synthetic materials that

contribute to environmental contamination [6][7]. The biocellulose is biodegradable biopolymer for friendly packaging, especially food protection. The usage of biocellulose as packaging has been limited due to the poor mechanical and barrier properties [6]. Biocellulose can perform as edible films for packaging food product and as carriers of foods additives such as antioxidant or antimicrobial agent to improve the characteristic of the edible films [8][9][10]. Edible films of biocellulose as food packaging containing antioxidants like catechin combined with bergamot essential oil can create a novel alternative to reduce oxidation in food products. The essential oils and catechin can demonstrate their ability in inhibiting free radical to increase food products with higher quality and safety. Previous study showed that methanol and water extract of *Citrus aurantium* L fruit revealed the antioxidant components. The total phenol content of the extracts ranged from 2.5 to 22.5 mg/g and 5.0 to 45.0 mg/g of pulp and peel fragments respectively

Corresponding Author: Yoice Srikandace, Indonesian Institute of Sciences-Research Unit for Clean Technology, Jl. Cicitu-Sangkuriang Bandung, Indonesia, yoice.srikandace.s@gmail.com

[11]. The major constituents of the different parts of *C. aurantium* L. essential oils were: β -pinene (0.62%–19.08%), limonene (0.53%–94.67%), trans- β -ocimene (3.11%–6.06%), linalool (0.76%–58.21%), and α -terpineol (0.13%–12.89%) [12][13][14]. The essential oils from leaves, flowers, fruits and peel of *Citrus aurantium*, *Citrus limetta* and *Citrus limon* served as the potential source of natural antioxidants [12][13][15]. The plants also produce secondary metabolite that perform antioxidant properties such as catechin [16]. Catechin is the natural phenol that belongs to flavonoid compound [17][18]. The total phenolic and total tannin contents of commercial black catechu and green tea, yellow tea, and black tea showed high antioxidant activities about 63.68-75.51% with IC₅₀ 3.92-3.31mg/mL [16][19]. The effectiveness edible film incorporated with catechin and essential oil is limited. There is no data that biocellulose incorporated with bergamot oil and catechin is studied. Therefore, the objectives of present study are to investigate antioxidant activity from edible films and to analysis the mechanical properties of edible film.

EXPERIMENT

Materials and preparation of bacterial cellulose-based edible film

Bergamot essential oil were purchased from Lansida Group. Nata de coco gels were purchased from local industry in Cianjur, West Java Province, Indonesia. Gels were washed and boiled with water containing NaOH 0.5% until the gels were stable on pH7. A total of 0.5 kg gel was blended with 200 mL water in order to obtain the slurry of biocellulose

Preparation of bacterial cellulose composites

An amount of 100 mL slurry was added with 1.0% carboxymethyl cellulose (CMC), 200 μ L glycerol, 0.1% Tween 80 and 0.5-2.5% essential oil and 0.005 of catechin. All composites was mixed in a beaker glass under magnetic stirring condition at 70°C, then poured on the trays. The trays were held overnight at 45°C in oven blower, then cooled at room temperature before peeling the films off the trays and stored in plastic bags until used.

Antioxidant assay

A total of 0.5 g of edible film was extracted with methanol for 1 x 24h to obtain methanol extract. For the evaluation, 2.0 mL of 0.1mM DPPH solution was added with 1.0mL of sample in a test tube, shaken and incubated in dark room for 30 minutes. Then, samples were read using a spectrophotometer at 517 nm. The antioxidant activity was calculated as percentage inhibition of DPPH, using the following equation:

$$\% \text{inhibition} = (\text{Ao} - \text{As}) / \text{Ao} \times 100 \%$$

Ao is the absorbance of DPPH and As is the absorbance of sample.

Mechanical properties

The mechanical properties of edible films were characterized by Orientec UCT-5T universal testing with 100 kgf load cell according to ISO 527- 1993E standard method. Dumbbell-shaped specimens were obtained from each film according to ISO 527-2 type 5A. The measurement was conducted at temperature of 23°C and relative humidity of 50%. At least five specimens of each sample was measured and computerized calculated to obtain the average value.

Fourier Transform Infra Red (FTIR) analysis

Fourier-transform infrared (FTIR) spectroscopy was conducted with Thermo Scientific Nicolet iS5 Spectrophotometer using the potassium bromide disk technique, in the range of 4000–500 cm⁻¹ with an attenuated reflectance (ATR).

Scanning Electron Microscope (SEM) analysis

Edible films were examined for surface characteristics using SEM JEOL JSM IT-300 JAPAN operating at 20kV. Film pieces were mounted on aluminum stubs using double-sided tape and then coated with a layer of gold (40–50 nm), allowed surface and cross-section visualization.

Gas chromatography mass spectrometry (GC-MS) analysis

The bergamot essential oil was analyzed using the Agilent 7890 B with MSD 5977A. One microliter of diluted sample (1/100 in methanol, v/v) was injected manually (split mode, split ratio 1:20). Calculation of peak area percentage was performed on basis of the FID signal using the GC HP-Chemstation software.

RESULT AND DISCUSSION

All edible films containing catechin (FC), Bergamot oil (FB), catechin and Bergamot oil (FCB) showed antioxidant activity and described in Table 1. Based on Table 1, the edible FC inhibited DPPH about 55.58-65.78% with 0.04-0.05% catechin concentrations. The film FB revealed percentage inhibition 67.08-73.16% with 2-2.5% bergamot oil. The edible FCB also presented percentage inhibition range in 50.17-81.07% with 1.0-2.5% oil concentrations as the best activity. Catechin has been known widely as antioxidant properties that isolated from tea plants. The peels of bergamot oil and *Citrus spp* produce aromatic and volatile compounds that perform antimicrobial and antioxidant activity. The combination of catechin and

bergamot oil increased inhibition rate against DPPH. It also indicated that the compounds were stable for edible films.

Table 1. Antioxidant activity of the edible films

Sample	Composite additives	Inhibition (%)
FC	0.01 % catechin	21.19
	0.02 % catechin	34.39
	0.03 % catechin	43.35
	0.04 % catechin	55.58
	0.05 % catechin	65.78
FB	0.5% oil	13.99
	1.0 % oil	33.13
	1.5 % oil	46.69
	2.0 % oil	67.08
	2.5 % oil	73.16
FCB	0.05 % catechin + 0.5 % oil	35.89
	0.05 % catechin + 1.0 % oil	50.17
	0.05 % catechin + 1.5 % oil	62.45
	0.05% catechin + 2.0 % oil	77.11
	0.05 % catechin + 2.5 % oil	81.07

Some studies showed the black catechu plants were rich source for catechin with percentage of free radical scavenging varied from 2.62-75.47% in black catechu and 74.18-77.06% in white catechu [16]. The Sichuan tea (green tea, yellow tea, and black tea) described the scavenging rate of hydroxyl radicals about 54.18 - 75.51% [19]. The study of *Citrus sinensis* and *C.aurantium* showed the antiradical activity varies from 58.48- 92.55% (leaves) and 55.46-88% (peels) [13][20][21].

Mechanical properties of all edible film were analyzed for tensile strength (TS) and elongation break (EB). The properties was described in Table 2.

Table 2. Mechanical properties of edible films

Sample	Tensile strength (Mpa)	Elongation (%)
Edible film A	62.72	20.00
FC	62.97	20.11
FB	58.12	22.50
FCB	58.27	22.31

Edible film A : film contained all composites (CMC, gliserol and Tween 80)

Table 2. showed the edible film A obtained 62.72Mpa (TS) and 20% (EB). The edible FC performed TS (62.97 Mpa) and EB (20.11%) did not show a significant different value of TS andEB from edible film A. On the

other side, edible films were enriched with bergamot oil (FB and FCB) decreased the TS value but increased the EB. The tensile strenght of FB and FCB decreased from 67.72 Mpa to 58.12-58.27 Mpa. Elongation of FB and FCB increased from 20.00% to 22.50-22.31%. The change of TS and EB value was due to interactions CMC and glycerol (polymer chains) with oil. The CMC, glycerol and oil filled, interfered the matrix of films and produced solid fine films. Glycerol also performed as plasticizer and demonstrated higher moisture content for biodegradable films [22]. The study of edible films with various concentrations of CMC, glycerol and some citrus oils showed range TS value 66-180 Mpa andEB value 22.9-11.8% as well[23][24]. It proved that CMC, glycerol and oils decreased the tensile forces but increased the elongation break.

The FTIR analysis (figure 1, figure 2, figure 3 and figure 4) described the characteristic bands of all films with addition of CMC, glycerol, catechin and oil.

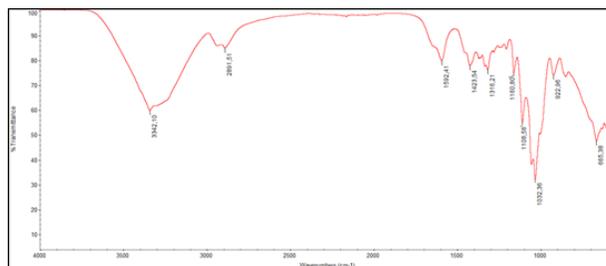


Fig 1 The FTIR spectra of biocellulose ,CMC and glycerol composite edible film.

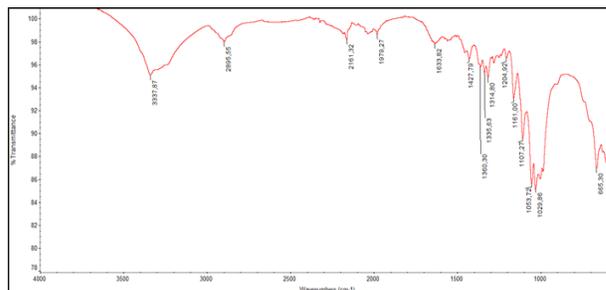


Fig 2 The FTIR spectra of biocellulose ,CMC, glycerol and catechin composite edible film.

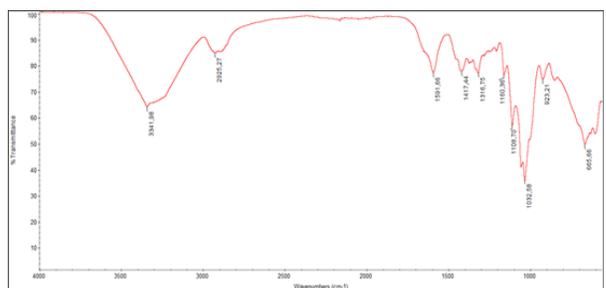


Fig 3 The FTIR spectra of biocellulose ,CMC, glycerol, catechin and 2% Bergamot oil composite edible film.

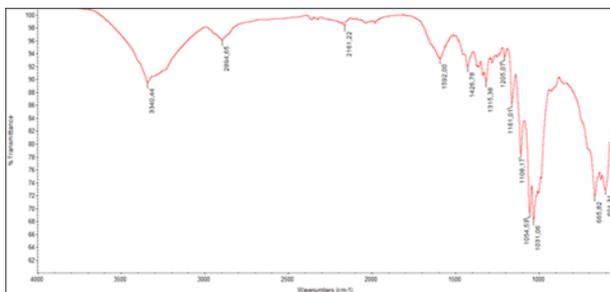


Fig 4 The FTIR spectra of biocellulose ,CMC, glycerol, catechin and 2% Bergamot oil composite edible film FBC

Based FTIR spectras, all films showed O – H group peak at 3337 – 3342 cm^{-1} , performed an important role in the physical properties of films. Peak at 2894-2925 cm^{-1} indicated the presence C-H group. The COO⁻ of CMC were obtained peak at 1592-1533 cm^{-1} . All spectra confirmed that the important bonds in cellulose polymer were present in the films even though the peaks and curves may change. Distinguish peaks of 3337 – 3342 cm^{-1} that should around 3400 cm^{-1} to 3500 cm^{-1} can be associated to the hydroxyl bonds of cellulose. The spectra region can be referred to as the intermolecular and intermolecular hydrogen bonds of cellulose [25][26].

Based the SEM analysis, edible film A (Fig 5) and FC (Fig 6) showed the surface of the films obtained cracks, roughness and many pores in the films because of the large molecule size of biocellulose. Composites were not dissolved with homogenous. The appearance of films containing bergamot oil (Fig. 7) and catechin-bergamot oil (Fig. 8) obtained the fibers more solid, smooth and homogenous. It was due to addition of essential oil. There was no remarkable structural difference between FB and FCB, so that Bergamot oil improved the flexibility and prevented cracks. Similar study, SEM analysis showed the surface microstructural of the edible film from breadfruit starch incorporated with essential oil of attarasa leaves was more rough and solid compared with film without incorporation of essential oils [27].

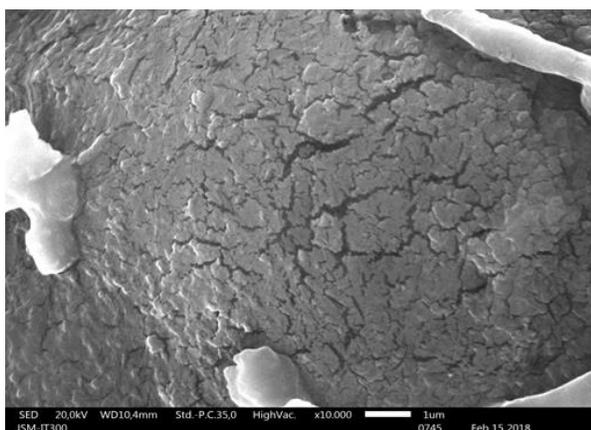


Fig 5 SEM micrographs of the surface of edible film A

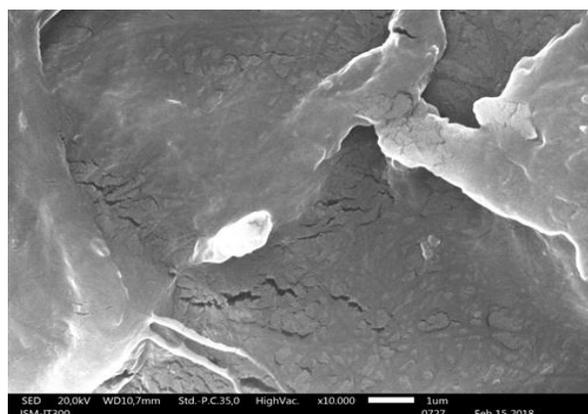


Fig 6 SEM micrographs of the surface of film FC

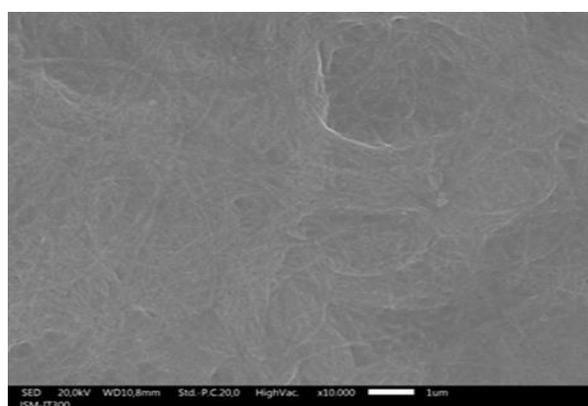


Fig 7 SEM micrographs of the surface of film FB

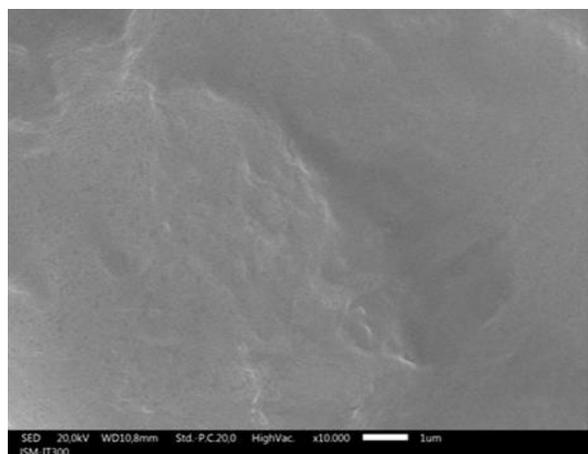


Fig 8 SEM micrographs of the surface of film FBC

The GCMS analysis results indicated the compounds of bergamot oil and described in Table 3 and Fig 9. Figure 9 and Table 3. showed bergamot oil contained 22 components accounting approximately 98.72% of the total bergamot oil contents.

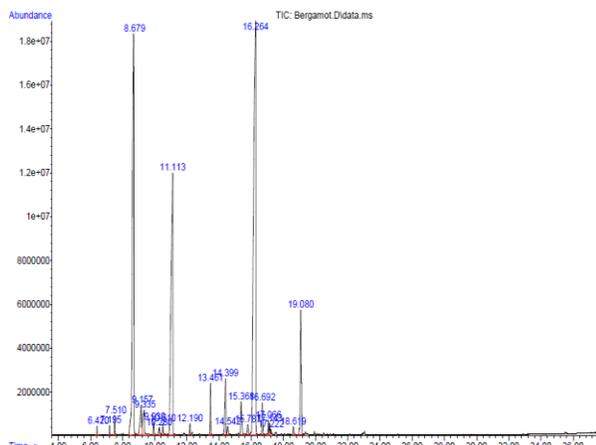


Fig 9 GC–MS analysis of volatile organic compounds of *Citrus aurantium*

The composition varied considerably, depending on the geographical origin but it also contained the typically compounds such as limonene, linalool and linalyl. Limonene is the principal component of *Citrus spp* oils (lime, lemon, sweet orange, and tangerine oils). A study revealed that the vacuum distillation of bergamot peels furnished a high-quality components of essential oil such as monoterpenes myrcene, linalool, and linalyl acetate [28]. The peels of *Citrus medica* L., *C. junos*, *C. limon* and *C. aurantifolia* contained the specific component such as camphene, citral and limonene [28][29].

Table 3 Composition of Bergamot oil

Compounds	Content (%)
α-pinene	0.16
β-phellandrene	0.19
β-myrcene	0.38
Limonene	23.66
2-Furanmethanol	0.40
Trans-linalool oxide (furanoid)	0.41
Linalool	19.44
Citral	1.01
Neral	0.47
Chloroacetic acid	1.44
α-terpineol	2.35
2,6-Octadien-1-ol	1.57
Linalyl acetate	37.88
α-Terpinyl acetate	5.31
Acetic acid	0.12
Cyclohexene	1.24
Pentafluoropropionate	0.25
Bornyl acetate	0.12
1,3-Cyclohexadiene	0.27
Cyclohexanol	0.30
3,5,5-Trimethylhexyl acetate	1.44
1,2-Dihydrolinalool	0.30

Citrus fruits have been a commercially important crop for thousands of years. Citrus essential oils are well known valuable in their flavor and fragrance properties, as well as numerous aromatherapeutic and medicinal applications. The essential oils will undoubtedly continue to play important roles in the food and beverage industries, especially for natural packaging.

CONCLUSION

Addition of catechin and bergamot oils into biocellulose based films showed an effect on mechanical properties. The film that contained bergamot oil presented the smooth surface compared to others and could enhance films' flexibility. The combination 0.05 % of catechin and 1-2.5% of bergamot oil showed the antioxidant activity with range inhibition 50.17-81.07 %.

ACKNOWLEDGMENTS

The authors acknowledge the financial support from INSINAS-LIPI, Indonesia.

REFERENCES

- [1] Alejandra, M.A.G., Sonia, H.C., Roberto, P.T.K. and Hafiz, M.N.I. 2016. Bacterial cellulose: A sustainable source to develop value-added products-A review. *Bioresources* 11, 1-15.
- [2] Gayathry, G. 2015. Production of Nata de Coco – a natural dietary fibre product from mature coconut water using *Gluconacetobacter xylinum* (sju-1). *Intl. J. Food. Ferment. Technol* 5, 231-235.
- [3] Darmawan, A.N. and Pradipta, A. 2015. Characterization of Nata de Coco Produced by Fermentation of Immobilized *Acetobacter xylinum*. *Agriculture and Agricultural Science Procedia* 3, 278 – 282.
- [4] Ali, A. and Nooshin, M.S. 2013. Bacterial Cellulose as High Performance Nano Biopolymer. *Research Journal of Pharmaceutical, Biological and Chemical Sciences* 5, 1324-1334.
- [5] Faezah, E., Norliza, A.R., Sahaid, M.K. and Siti, M.T. 2017. Effect of agitation conditions on bacterial cellulose production by *Acetobacter xylinum* 01416 in fermentation of matured coconut water medium. *Malaysian Journal of Analytical Sciences* 21, 261-266.
- [6] Ana R.V.F., Vítor D.A. and Isabel M.C. 2016. Review: Polysaccharide-based membrane in food packaging applications. *Membranes* 6, 1-17.
- [7] Henriette M.C.A., Morsyleide, F.R. and Luiz H.C.M. 2016. Nanocellulose in bio-based food packaging applications. *Industrial Crops and Products* 1, 1-8.
- [8] Véronique C. 2013. Polysaccharide-based biomaterials with antimicrobial and antioxidant properties. *Polimeros* 23, 287-297.

- [9] Bhanu, M., Anu, K. and Harsha, K. 2015. Antimicrobial food packaging: potential and pitfalls. *Frontiers in microbiology* 6, 1-9.
- [10] Reyhan, I. and Ozlem, K.E. 2015. Novel food packaging systems with natural antimicrobial agents. *J Food Sci Technol* 52, 6095–6111.
- [11] Prakash, J.Divya., Prakash, J. and Lakshmi A.J. 2016. Antioxidant properties of fresh and processes *Citrus aurantium* fruit. *Cogent Food & Agriculture* 2, 1-12.
- [12] Eirini, S., Paschalina C., Kortessa, D.T. and Ioannis, T. 2013. Volatile constituents and antioxidant activity of peels, flowers and leaf oils *Citrus aurantium* L growing in greece. *Molecules* 18, 10639-10647.
- [13] Majnooni, M., Kamran, M., Gholivand, M., Ali, M., Hamid, R.M.M., Afnanzade, N., Abolghasemi, M. and Marzieh, P. 2015. Chemical composition, cytotoxicity, antioxidant activities of essential oil from the leaves of *Citrus aurantium* L. *African Journal of Biotechnology* 11, 498-503.
- [14] Djamel, D. 2015. Chemical profile, antibacterial and antioxidant activity of Algerian citrus essential oils and their application in *Sardina pilchardus*. *Foods* 4, 208-228.
- [15] Muthiah, P.L., Umamaheswari, M., and Asokkumar, K. 2012. In vitro antioxidant activities of leaves, fruits and peel extracts of citrus. *International Journal of Phytopharmacy* 2, 13-20.
- [16] Thidarat, D., Chanida, P. and Nijsiri, R. 2014. Catechin and epicatechin contents and antioxidant activity of commercial black catechu and pale catechu. *Journal of Chemical and Pharmaceutical Research* 6, 2225-2232.
- [17] Michalina, G., Katarzyna, N., Grzegorz, B. And Isabella, S. 2017. Antioxidant properties of catechins: comparison with other antioxidants. *Food chemistry* 1, 1-49.
- [18] Jurga, B. and Dalia M.K. 2018. The role of catechins in cellular responses to oxidative stress. *Molecules* 23, 1-11.
- [19] Jianhua, L., Shengxiang, C., Mingzhu, Z. and Xueli, M. 2017. Comparison of catechins and antioxidant activity in four kinds of Shicuan tea. IOP Conf. Series: *Earth and Environmental Science* 94, 1-6.
- [20] Samira, L. and Khodir, M. 2013. Phenolic contents and antioxidant activity of orange varieties (*Citrus sinensis* L. and *Citrus aurantium* L.) cultivated in Algeria: Peels and leaves. *Industrial Crops and Products* 50, 723-730.
- [21] Mila R., Antonija, P. and Franko, B. 2018. Chemical Composition and Antioxidant Activity of Essential Oil Obtained from Bitter Orange Peel (*Citrus aurantium* L.) Using Two Methods. *Croat. Chem. Acta* 91, 125–128.
- [22] Sanyang, M.L., Sapuan, S.M., Jawaid, M., Ishak, M.R. and Sahari, J. 2009. Effect of plasticizer type and concentration on physical properties on biodegradable films based on sugar palm (*Arenga pinnata*) starch for food packaging. *J Food Sci Technol* 53, 326-336.
- [23] Lucia, I., Indriyati, Anung, S. and Sri, P. 2016. Physical and mechanical properties of modified bacterial cellulose composite films. *AIP Conference Proceedings* 1711, 1-5.
- [24] Lucia, I. and Indriyati. 2017. Incorporation citrus essential oils into bacterial cellulose based edible films and assessment for their physical properties. *IOP.Conf.Series: Earth and Environmental Sciences* 60, 1-5.
- [25] Adebayo, B.C., Akintunde, M.O. and Alao, S.O. 2017. Comparative Effect of Agrowastes on Bacterial Cellulose Production by *Acinetobacter* sp. BAN1 and *Acetobacter pasteurianus* PW1. *Turkish Journal of Agriculture and Natural Sciences* 4, 145-154.
- [26] Emilia, L.B., Concetta, C., Maria, L.D., Antonella, L., Angelo, L., Francesca, P., Carlo S. and Maria, C.V. 2008. Comparison of the volatile constituents in cold-press of Bergamot oil and a volatile oil isolated by vacuum distillation. *J. Agric. Food Chem.* 55, 7847–7851.
- [27] Cut, F.Z., Jamaran, K., Marpongahtun and Erman M. 2013. Effect of essential oil of Attarasa leaf (*Litsea cubeba* Lour.Pers) on Physico-mechanical and microstructural properties of breadfruit starch-alginate edible film. *Malaysian Journal of Analytical Sciences* 17, 370 - 375.
- [28] Aliyah, A.H., Rante, H. and Mufidah, D.RN. 2017. GC-MS analysis and antimicrobial activity determination of *Citrus medica* L var. proper leaf from South Sulawesi against skin pathogen microorganism. *IOP Conf. Series: Materials Science and Engineering* 259, 1-6.
- [29] Joon, H.H., Naeem, K., Nargis, J. Young, S.H., Eun, Y., Ji, Y.C., Cheong, M.L. and Kyong, S.K. 2016. Review: Determination of Volatile Flavour Profiles of *Citrus* spp. Fruits by SDE-GC–MS and Enantiomeric Composition of Chiral Compounds by MDGC–MS. *Phytochemical analysis* 1, 1-12.